

## Survey of *Aeromonas hydrophila* in three marine fish species from north west Arabian Gulf , Iraq

Ghazi M. Al-Maleky

Rajaa A. Hanef

Department of Marine Biology- Marine Science Center-University of Basrah- Iraq.

### Abstract

This study preformed on 74 samples of marine water fishes (24 of *Epinephelus tauvina*, 27 of *Hilsa ilisha* and 23 of *Lethriuns nebulosus*). Which collected from North West Arabian Gulf of Basrah. All samples were examined for the presence of *Aeromonas hydrophila* in muscles. 24 isolates of *Aeromonas hydrophila* were obtained, 8 (33.3%) from *Epinephelus tauvina*, 12 (44.4%) from *Hilsa ilisha*, and 4 (17.39%) from *Lethriuns nebulosus*. Then all isolates were examined for their ability to hemolytic activity as a virulence factor, the higher percentage of hemolytic activity isolates was found in *Epinephelus tauvina*.

### مسح جرثومة *Aeromonas hydrophila* في ثلاثة انواع من الاسماك البحرية من شمال غرب الخليج العربي

رجاء عبد الكاظم حنف

غازي مالخ جابر

قسم الاحياء البحرية- مركز علوم البحار- جامعة البصرة

### المستخلص

أجريت هذه الدراسة على 74 عينة من اسماك المياه البحرية (24 من *Epinephelus tauvina* و 27 من *Hilsa ilisha* و 23 من *Aeromonas hydrophila* في عضلات الانواع الثلاثة من الاسماك. وظهرت النتائج عن وجود 24 عذلة من الجرثومة منها 8 (33.3%) في *Epinephelus tauvina* و 12 (44.4%) في *Hilsa ilisha* و 4 (17.39%) في *Cyanoglossus arel*. كما تم اختيار العزلات إلى قابليتها على تحليل الدم كعامل ضراوة, وبينت الدراسة إن العذلة التي حققت أعلى نسبة مئوية للضراوة تم الحصول عليها من *Epinephelus tauvina*.

## Introduction

Fish is a very perishable, high-protein food that typically contains a high level of free amino acids and volatile nitrogen bases which are essential for human consumption, in addition to high polyunsaturated fatty acids (González-Fandos *et al.*, 2005).

The bacterium *Aeromonas* is considered as one of the newly emerging water and food born pathogens (Merino *et al.*, 1998; Gugnani, 1999). In fish *Aeromonas* typically causes hemorrhagic septicemia and has been implicated in different outbreaks associated with heavy losses (Son *et al.*, 1997). Isolation of these organisms has been reported from a variety of food including fishes (Adiththepchaikram *et al.*, 2008).

Members of the genus *Aeromonas* are Gram-negative non spore-forming straight rods, which occur singly, in pairs or short chains. They are facultative anaerobes, being both catalase and oxidase positive. The aeromonads break down carbohydrates with the production of acid or acid and gas. Most of the mesophilic species within this genus are motile and have a single polar flagellum. Swarming motility with the production of lateral flagella has also recently been described (Kirov *et al.*, 2002; Andrade *et al.*, 2006). Although strains of *A. salmonicida* are capable of producing lateral flagella they are non-motile. This thought to be as a result of inactivation of the *lafA* (flagellin gene) by transposase 8 (IS3 family) (Merino *et al.*, 2003). This bacteria is widely distributed in an aquatic environment (Fiorentini *et al.*, 1998) this bacteria infect humans and causing septicemia, gastroenteritis, acute diarrhea urinary tract infection and ear infection (Koneman *et al.*, 1994; Topic *et al.*, 2000).

*Aeromonas* group produce number of potential virulence factors, including, enterotoxins, haemolysins, cytotoxins and proteases (Burke *et al.*, 1982; Ljungh and Wadström, 1983) mentioned that the hemolytic activity is strongly associated with enterotoxin production in members of *Aeromonas* genus. Rogulska *et al.*, (1994) reported that the hemolytic activity of *Aeromonas* species act as marker of pathogenicity. The mechanism of

action of the enterotoxins is similar to that of *Vibrio cholera* (Tanoue *et al.*, 2005).

The aeromonads can grow at range of temperature from 5 to 44 °C. The optimum temperature for growth is 22- 28 °C. Most isolates of clinical significance will grow readily at 37 °C. The pH range for growth is 5.5 -9.0 growth is inhibited in 6% salt broth. The G and C content of DNA is 57-63%. The aim of the study for detection of the presence of *Aeromonas hydrophila* in marine fishes and its association with human health.

## Materials and Methods

### **Isolation and identification:**

74 samples of Marine fishes (24, *Epinephelus tauvina*; 27, *Hilsa ilisha*; 23, *Lethrinus nebulosus*) were collected from marine water of north west Arabian Gulf 29 40 787 N 48 43 750 E (Awama 1), then samples were transferred to the laboratory under septic condition for bacteriological examination.

Enrichment method used for analyses the samples according to Okrend *et al.*, (1987). 12.5 gm of muscle tissue from each sample was added to 112.5 ml of trypticase soybroth containing 5 mg ampicillin /ml and blended for 2 minutes, then diluted up to 10<sup>-3</sup> in buffered phosphate diluent, and the count was carried out by aforementioned dilution as recommended by Palumbo *et al.*, (1989) using Macconkey manitol ampicillin agar. The number of colonies which showed red color in countable plates was enumerated as *Aeromonas* organism and diagnosis was confirmed by biochemical tests.

### **Hemolytic activity test**

Determination of hemolytic activity of the isolated strains it was carried out using 5% sheep blood agar as recommended by Rogulska *et al.*, (1994).

### **Biochemical tests**

Biochemical tests such as gram stain, Motility, Indole, Voges proskauer, Methyl red,

Urase, H<sub>2</sub>S, Nitrate reduction, Catalase, Oxidase, Glucose, Maltose, Sucrose, Hemolysis, Gelatin liquification, Ornithine decarboxylase and NaCl tolerance were used for diagnosis of *Aeromonas hydrophila*. (Kirov *et al*, 2002).

### Results and Discussion

This study was performed to detect the presence of *Aeromonas hydrophila* in marine

fishes, Biochemical tests were used for diagnosis of *Aeromonas hydrophila* as showing in table (1). The results in table (2) was showed that 8(33.3%), 12(44.4%), 4 (17.39%) isolates were obtained from *Epinephelus tauvina*, *Hilsa ilisha* and *Lethrius nebulosus* respectively. Figure 1 shows the colonies of *Aeromonas hydrophila*, Figure 2 shows infection in A *Epinephelus tauvina*, B *Hilsa ilisha* and C *Lethrius nebulosus*.

Table 1: Morphological and biochemical characteristics of *Aeromonas hydrophila* isolated from marine fishes

Sr.	Characteristic	<i>Aeromonase hydrophila</i> isolated
1.	Gram stains	-
2.	Shape	Rod
3.	Motility	M
4.	Indole test	+
5.	Voges proskeur test	-
6.	Methyl red test	+
7.	Urase test	-
8.	H <sub>2</sub> S gas	-
9.	Nitrate reduction test	+
10.	Catalase test	+
11.	Oxidase test	+
12.	Glucose	+
13.	Maltose	+
14.	Sucrose	-
15.	Hemolysis	β
16.	Gelatin liquification	+
17.	Ornithine decarboxylase	+
18.	NaCl 0%	+
19.	NaCl 6%	+

Table 2: Ocurance of *Aeromonas hydrophila* in three marine Fish species

Marine fishes	No. of samples	No. of isolates	Percentage %
<i>Epinephelus tauvina</i>	24	8	33.3
<i>Hilsa ilisha</i>	27	12	44.4
<i>Lethrius nebulosus</i>	23	4	17.39
Total	74	24	95.09

The present results disagree, with those reported by Okrend *et al.*, (1987); Palumbo *et al.*, (1989) and Freitas *et al.*, (1992) since the authors pointed out that hemolysin was detected in 100% of *Aeromonas hydrophila* strains recovered from some varieties of food.

Abyta *et al.*, (1994) identified *Aeromonas hydrophila* as the primary enterophogenic species, In addition, beta hemolytic strains of *Aeromonas* are assigned to *Aeromonas hydrophila* (Deodhor *et al.*, 1991), Varnam and Evanus, (1991) reported that a number of phenotypic characters have been proposed as a markers of enteropathogenicity of *Aeromonas* species and they added that the most

important of these markers was hemolysin production.

The information given by the achieved results reveled that *Aeromonas* organism existed in the examined fishes and therefore the foods may play a significant role in the epidemiology of gastroenteritis for human, so the strict hygienic measures, good food handling practice at home, properly clean, sanitary equipments and contact surfaces should be recommended to avoid contamination with *Aeromonas* organism. *Aeromonas hydrophila* consider as etiologic agent for tail/fin diseases and hemorrhagic septicemia of fresh water fishes.



Fig (1): colonies of *Aeromonas hydrophila* on mackonkey manitol ampicilin agar.



A



B



C

Fig. (2): infection in A: *Epinephelus tauvina*, B: *Hilsa ilisha* and C: *Lethrius nebulosus*.

## References

- Abyta, C.; Palumbo, S. A. and Stelma, G. N. (1994): *Aeromonas hydrophila* group, Ch. I in Y. H. Hui; J. r. Garham, K.D. Murrell and D. O. Cliver (ed), food borne disease handbook. Diseases caused by bacteria. Marcel Dekker, Inc., New york.
- Adithepchaikarm, P.; Parichat, P. and Pongask, R. (2008): potential of psidium ganjava supplemental fish diets in controlling *Aeromonas hydrophila* infection in Tilapia (*oreochromis niloticus*) J. of Bioscience and Bioengineering. 106, 5: 224 – 230.
- Andrade, J. M.; Cairrao, F. and Arraino, C. M. (2006): RNase R affects gene exp. Regulation of omp A. I, Mol. Microbial. 60: 219-228.(pabmed)
- Burke, V.; Robinson, J.; Atkinson, H. M. and Gracey, M. (1982): Biochemical characteristics of enterotoxigenic *Aeromonas* spp. Journal of Clinical Microbiology. 15(1): 48-52.
- Deodehar, L.P.; Saraswath, K. and Varudkar, V. (1991): *Aeromonas* spp. And their association with human diarrhea disease. J. Clin. Microbiol., 29: 853-856.
- Fiorentini, C.; Barbieri, E.; Falzano, L.; Matrese, P.; Baffpne, W.; Pianetti, A.; katouli, M.; kinhu, I.; Miolby, R. R.; Bruscolini, F.; Casiere, A. and Donelli, G. (1998): Ocurance , diversity and pathogencity of mesophitic *Aeromonas* in estuarine water of Italian coast of the Adriatic Sea Appl. Microbial. 85 (3) ; 501 – 511.
- Freitas, A. C.; Nunes, M. P.; Milhomemn, A. M. and Ricciardi, I. D. (1992): Occurrence of species in pasteurized milk and white cheese Reo de Janeiro, Brazil. J. Food. Prot., 56: 62-65.
- González-Fandos, E.; Villarino-Rodríguezb, a. A.; García-Linaresb, M. C.; García-Ariasb, M. T. and García-Fernándezb, M. C. (2005): Microbiological safety and sensory characteristics of salmon slices processed by the sous vide method. Food Control. (16) 1: 77-85.
- Gugnani, H. C. (1999): Some emerging food and water borne pathogens. Commun. Dis. 31(2):65-72.
- Kirov, S.M.,Tassell, Semmler, A. B. T. ( 2002). Lateral flagella and swarming motility in *Aeromonas* species. J Bacterial, 184, 547-555.
- Koneman, E. W.; Allen, S. D.; Janda, W. M. Sihreckenberger, P.C.and Winn, W. C. Jr.(1994): Introduction to Diagnostic Microbiology J. B. Lippinctt Company PP. 117 – 123.
- Ljungh, A. and Wadström, T. (1983): Toxins of *Vibrio Parahaemolyticus* and *Aeromonas hydrophila* J. Toxicol. Toxin Rev.1: 257-259.
- Merino, S., Gavin, R., Vilches, S. (2003). Acolinization factor (production of lateral flagella of mesophilic *Aeromonase* spp, is inactive in *Aeromonase salmonicida* strains. Appl. Environ. Microbrol, 69, 663-667.
- Merino, S.; Noguerras, M. M.; Aguilar, A.; Rubires, X.; Albertí, S.; Benedí, V. J. and Tomás, J. M. (1998): Activation of the Complement Classical Pathway (C1q Binding) by Mesophilic *Aeromonas hydrophila* Outer Membrane Protein. Infect Immun. 66(8): 3825-3831.
- Okrend, J. G. A.; Rose, B. B. and Bennett, B. C. (1987): Incidence and Toxygenicity of *Aeromonase* species in retail poultry, beef and pork. J. Food Port. , 50: 509 – 513.
- Palumbo, S. A.; Bencivengo, M. M.; Corral, F. D.; Williams, A. C. and Buchanan, R. I. (1989):

Characterization of *Aeromonas Hydrophila* group isolated from retail food of animal origin . J. Clin. Microbial., 27: 824 – 829.

Rogulska, A.; Antychowicz, J. and Żelazny, J. (1994): Aktywność hemolityczna i proteolityczna *Aeromonas hydrophila* i *Aeromonas sobria* dla karpia (*Cyprinus carpio*) [Hemolytic and proteolytic activity as an indicator of pathogenicity of the bacteria *Aeromonas hydrophila* and *Aeromonas sobria* for carp (*Cyprinus carpio*)]. Med. Wet. 50: 55-58.

Son, R.; Rusul, G.; Sahilah, A. M.; Zainuri, A.; Raha, A. R. and Salmah, I. (1997): Antibiotic resistance and Plasmid profile of *Aeromonas Hydrophila* isolated from cultured fish (*Tilapia mossambica*). Appl. Microbial, 24 (6) 479 – 482.

Tanoue, N., Takahashe , A., Okamoto, K. et al (2005). Apore-forming toxin produced by *Aeromonas sobria* activates CAMP-dependent Cl (-) secretory pathways to cause diarrhoea. FEMS Microbiol Lett, 242, 195-201.

Topic, I. N.; Teskeredgic, E. L.; Ctrunjak, P. and Rakovac, Co 7. (2000): *Aeromonas hydrophila* isolated wild fresh water fish in Croatia – V. Research communications, 24, 371 – 377.

Varnam, A. H. and Evans, M. G. (1991): *Aeromonas* in foodborne pathogen. Wolfe publishing ltd. London.